REVIEW



What microRNAs could tell us about the human X chromosome

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Abstract

MicroRNAs (miRNA) are small-non coding RNAs endowed with great regulatory power, thus playing key roles not only in almost all physiological pathways, but also in the pathogenesis of several diseases. Surprisingly, genomic distribution analysis revealed the highest density of miRNA sequences on the X chromosome; this evolutionary conserved mammalian feature equips females with a larger miRNA machinery than males. However, miRNAs contribution to some X-related conditions, properties or functions is still poorly explored. With the aim to support and focus research in the field, this review analyzes the literature and databases about X-linked miRNAs, trying to understand how miRNAs could contribute to emerging gender-biased functions and pathological mechanisms, such as immunity and cancer. A fine map of miRNA sequences on the X chromosome is reported, and their known functions are discussed; in addition, bioinformatics functional analyses of the whole X-linked miRNA targetome (predicted and validated) were performed. The emerging scenario points to different gaps in the knowledge that should be filled with future experimental investigations, also in terms of possible implications and pathological perspectives for X chromosome aneuploidy syndromes, such as Turner and Klinefelter syndromes.

Keywords microRNA · X chromosome · Turner syndrome · Klinefelter syndrome

Introduction

MicroRNAs (miRNA) are small non-coding RNAs that post-transcriptionally regulate gene expression by affecting both translation and stability of complementary mRNAs [1]. Bio-informatics predictions indicate that mammalian miRNAs regulate 30–50% of all protein-coding genes; each miRNA

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can bind several mRNAs and each mRNA can be targeted by different miRNAs, thus giving rise to complex regulatory networks that take part in the regulation of almost all physiological pathways [2, 3]. As a consequence, miRNA mutations, dysregulation of their expression or dysfunction of miRNA biogenesis have a key role in different pathological processes, with oncogenesis as the most investigated field [4–6].

miRNA genes can be present singularly in the genome or clustered together, with their own promoter or hosted by coding or non-coding genes, thus generally sharing their transcription promoter [7]. They are transcribed by RNA polymerase II as long primary transcripts (pri-miRNAs), wherein miRNA sequences fold into hairpin structures, recognized and excised by Drosha and DGCR8, the microprocessor complex, generating 60–80 nt precursors (pre-miRNAs). Pre-miRNAs are exported to the cytoplasm where they are processed by Dicer in miRNA duplexes. Finally, the mature miRNA strand that could derive from the 5' arm or 3' arm of the pre-miRNA (miRNA-5p or miRNA-3p, respectively) is loaded onto an Argonaute protein within the RISC complex to bind and silence complementary mRNA targets [8].



Surprisingly, genomic distribution analysis revealed that the human X chromosome has a higher density of miRNAs when compared to autosomes [9]; in contrast, the Y chromosome has only 4 miRNA sequences, but not experimentally validated, with 2 produced by the PAR1 (pseudoautosomal region 1) shared by both sex chromosomes. The miRNA enrichment on the X chromosome was observed in several mammalian species, but does not extend to species other than mammals [9]. The evolutionary conserved higher density of miRNAs on the X chromosome may suggest that X-linked miRNAs could contribute to some X-related conditions, properties or functions in mammals.

It has long been known that the presence of two X chromosomes in females and only one X chromosome in males requires mechanisms to equalize gene dosage between sexes and relative to autosomes to avoid a potentially lethal double-dose of genes residing on the X chromosome [10]. This mechanism involves two processes: X chromosome inactivation (XCI), i.e., the silencing of almost all genes on the one X chromosome leading to partial functional X monosomy, and X chromosome upregulation leading to increased gene expression on the single active X chromosome in males or females. It has been shown that up to 15% of the X-linked genes escape permanent silencing ("escapees"), with large variability in their number and tissue distribution within a given individual and between individuals: the escape from silencing or skewed XCI allow the expression of some genes by both X chromosomes in females [11, 12]. Furthermore, during early female embryonic development, the process of XCI is random across alleles in all cells, i.e., occurs irrespective of the parental origin of X chromosome and is clonally maintained once established, thus resulting in females being a functional mosaic for the active X chromosome across cell types. The XCI is a multistep process, involving a mechanism of counting and choice of the chromosome that will start XCI. The entire course of action is directed by the X inactivation center (Xic), a nuclear complex constituted by many non-coding DNA elements and genes (XIST, etc.) that lead to packaging into transcriptional inactive heterochromatin [13], also known as the Barr body.

X chromosome aneuploidies interfering with dosage compensation cause widely variable symptoms in two different syndromes, in particular Turner syndrome (with the classical karyotype 45,X), caused by a partial or complete lack of a second X chromosome in females, and Klinefelter syndrome

(with the classical karyotype 47,XXY), caused by two or more X chromosomes in males. In both syndromes no obvious genotype-phenotype relationship has so far been established, and patients carrying the same karyotype may exhibit widely differing traits, suggesting a role of epigenetic mechanisms behind sex chromosome aneuploidy [14, 15]. So far, some studies have been dedicated to this issue, essentially based on the analysis of epigenetic mechanisms, as DNA methylation and histone modifications [16, 17], and recently a dose effect for the number of X chromosomes has been shown to affect the expression profile for a range of mRNAs in a comparison of 45,X, 46,XX, 46,XY and 47,XXY individuals [18], implying that a similar dose effect could be present for X-linked miRNAs. But hitherto, the contribution of miRNAs to the phenotype of these syndromes has very poorly investigated [19–21], even though the higher density of miRNAs mapped on the X chromosome and their recognized regulatory role in biological processes.

In this review, we analyze the literature and databases about X-linked miRNAs, aiming at understanding how miRNAs could contribute to emerging gender-biased functions and to highlight some gaps in the knowledge, particularly in terms of possible implications and pathological perspectives for X chromosome aneuploidy syndromes.

Mapping of miRNA sequences on X chromosome

A general overview

The human X chromosome contains 10% of all miR-NAs detected so far in the human genome. According to the miRbase database, 118 miRNAs are located on the X chromosome, with 62 classified as "with confidence", i.e., experimentally validated. Exploring Ensembl and miRbase databases (https://www.ensembl.org/Homo_sapiens/; https://www.miRBase.org), positions of the miRNA sequences were mapped on the X chromosome as well as their genomic context, i.e., their possible position in host gene and transcribed strand. The full miRNA list is available as Supplementary Material, whereas some statistical parameters are reported in Table 1: approximately 70% of miRNA sequences are localized on Xq, the phylogenetically oldest part of the X chromosome [22]; more than the half (62.7%)

Table 1 Information about miRNA sequence mapping on X chromosome, based on Ensembl and miRBase databases

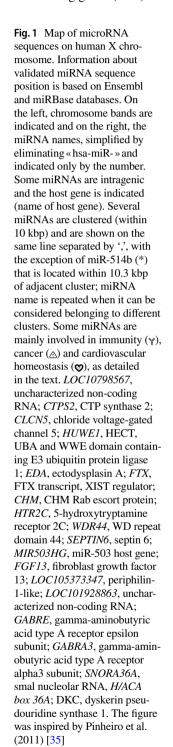
| miRNAs | Xp | Xq | Strand | Intragenic (same strand of host gene) | Intergenic |
|---------------------|----|----|----------------|---------------------------------------|------------|
| 62 "Confidence" | 14 | 48 | 16 (+), 46 (-) | 41 (31) | 21 |
| 56 "Non confidence" | 22 | 34 | 25 (+), 31 (-) | 33 (24) | 23 |
| 118 Total | 36 | 82 | 41 (+), 77 (-) | 74 (55) | 44 |

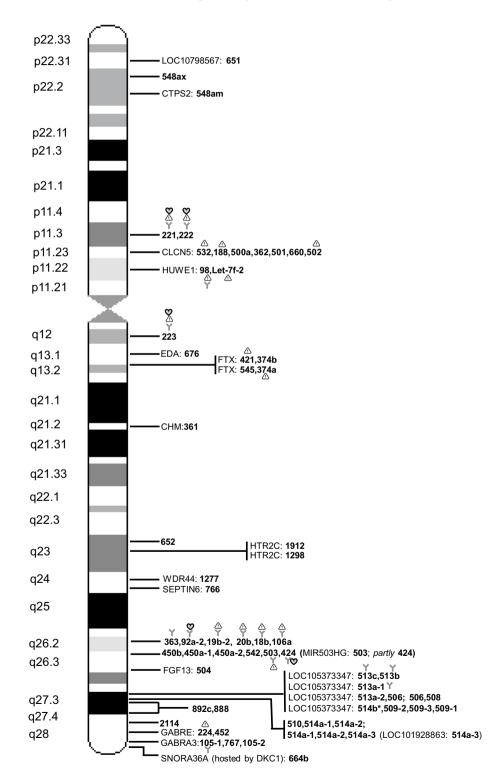


are intragenic, suggesting a co-regulation of miRNAs and host genes, in particular when they are transcribed by the same strand of host gene, and indeed this is the most often the case. We focused our attention on the 62 miRNAs experimentally validated and they are reported in Fig. 1; most of them are clustered (77.3%), and hosted in either coding or non-coding genes (66%).

Genes related to X-linked syndromes: any relationships with hosted miRNAs?

Some of the hosting coding genes are directly involved in X-linked syndromes. This is the case of *HUWE1*, an E3 ubiquitin ligase required for the ubiquitination and subsequent degradation of different targets, such as the







anti-apoptotic protein Mcl1 [myeloid cell leukemia sequence 1 (BCL2-related)], the tumor suppressor p53, core histones and DNA polymerase beta; mutations in the HUWE1 gene are associated with X-linked syndromic cognitive disability, where researchers found copy number variations [23], that perhaps could have similarities with some of the neurocognitive challenges seen in some individuals with Turner syndrome. It has also been proposed that an increased propensity to harbor copy number variations should be present in Turner syndrome (45,X) [24], while no studies so far have investigated this in the other sex chromosome aneuploidy syndromes. Defects in the EDA gene, hosting miR-676 and encoding a type II membrane protein belonging to the tumor necrosis factor family, are a cause of ectodermal dysplasia, anhidrotic, which is also known as X-linked hypohidrotic ectodermal dysplasia [25]. Another hosting miRNA gene, CHM, has been involved in an X-linked disease, choroideremia, also known as tapetochoroidal dystrophy, characterized by progressive dystrophy of the choroid, retinal pigment epithelium and retina [26]. Possible miRNA contribution to all those syndromes involving X-linked genes hosting miR-NAs has not been explored.

Noteworthy, two different miRNA clusters are also hosted (and likely co-regulated) by the FTX gene, located upstream of XIST within the X-inactivation center (XIC), and producing a long non-coding RNA endowed with a central role in positively regulating the expression of XIST, essential for the initiation and spread of X-inactivation [27]. A relationship between FTX and miR-374a has been reported: FTX sponges miR-374a, resulting in the repression of Wnt/β-catenin signaling activity and consequent inhibition of hepatocellular carcinoma cell (HCC) epithelial-mesenchymal transition and invasion; moreover, FTX inhibits proliferation of HCC cells by binding the DNA replication licensing factor MCM2, and it is significantly downregulated in HCC tissues compared with normal liver tissues. These features point to a tumor suppressor role for FTX: together with the notion that FTX is expressed at higher levels in female in comparison to male livers, it could be speculated that the described mechanisms contribute to the higher susceptibility of males than females to HCC [28].

Finally, although a precise map of XCI escapee miRNAs is still missing, some miRNAs are located within genes that have been shown to escape XCI, i.e., CTPS2, HTR2C, GABRE [29], FTX [27], suggesting that also the hosted miRNAs could escape silencing. It is known that with female mosaicism, i.e., random inactivation of one of the two parental X chromosome and clonal transmission to the daughter cells, silencing escapees or skewed patterns of inactivation of X genes could lead to unbalanced expression between sexes and to sex-specific responses; however, miRNA contribution to these phenomena has not been explored, even though the existence of multiple targets for each miRNA

could result in a cascade-like effect, leading to greater gender differences in terms of gene regulation.

Role of X-linked miRNAs emerging in the literature

Immunity and cancer

Based on literature data, the two emerging (or most studied) pathways involving regulation by X-linked miRNAs are immunity and cancer, two processes strictly linked with each other.

A significant difference between the male and female immune system function has been described: from childhood to old age, the female immune system appears more flexible, appropriate and able to better counteract infections and non-infectious disease, including cancer; females have longer life expectancies and an improved survival outcome from shock episodes caused by sepsis, injury or trauma-hemorrhage. However, this advantage can become detrimental, since the sex-biased nature of the immune system can result in an increased susceptibility to develop autoimmune diseases [30, 31]. Gender-specific immune responses may be partially explained by hormonal regulation; however, the importance of the X chromosome in shaping immune functions and autoimmunity has been recognized, since the presence of several X-located genes with a direct or an indirect role in immunity, including several members of the Toll-like receptor family pathway, with some of them being responsible for X-linked primary immunodeficiency (PID) [32-34]. A number of X-linked miRNAs impacting the immune system integrity and function potentiate the concept of the immunological advantage of females [reviewed in 35]. Among the X-linked miRNAs involved in immune regulation, miR-223 is probably the most studied so far. miR-223, which is also involved in cancer pathology, is expressed in the myeloid lineage in the bone marrow, and is a regulator of neutrophil differentiation from myeloid precursors; it is also a negative modulator of the inflammatory response [36, 37]. Other miRNAs mapping on X chromosome are involved in hematopoietic lineage differentiation. miR-106 was shown to negatively control monocytopoiesis by targeting AML-1; intriguingly, all miRNAs of the miR-106-363 cluster have oncogenic potential and been implicated in human T cell leukemias, where they are overexpressed [38]. In addition, two clustered miRNAs, miR-424 and miR-503, together with miR-222, all X-linked, promote monocytic differentiation [39]. The involvement of X-linked miRNAs in gender-biased immunity is also supported by a study reporting differential expression of six X-linked miR-NAs (miR-221, miR-222, miR-98, miR-532, miR-106 and



miR-92a) in PBMC (peripheral blood mononuclear cells) between males and females affected by rheumatoid arthritis, an autoimmune disease that affects females three times more often than men [40].

An example of a miRNA-dependent, sex-specific regulation of immune responses and cancer immunosurveillance is represented by the PD-1/PD-L1 pathway. The PD1 receptor, by binding to PD-L1 (or PD-L2), induces T cells to undergo immunosurveillance against tumors; PD-1 is also considered an immune checkpoint key molecule. Intriguingly, a number of X-linked miRNAs directly targets the stability and/or the translation of PD-L1 transcript (i.e., miR-20a/b, miR-106 a/b, miR-513, miR-424), or interfere with the transcription factors modulating its expression, i.e., miR-18a/b targeting HIF and miR-221/222 targeting STAT3 [31].

Several forms of cancer show a gender difference in terms of incidence, prevalence, and response to therapy [41]. miR-221 and miR-222 are the most extensively studied in tumors of different origins, wherein they act as oncomirs (oncogenic miRNAs) controlling the development and progression of the tumor through the down-regulation of several key targets [42, 43]. Both miRNAs are located in a cluster on the X chromosome; their deregulation is a hallmark of several types of cancer, probably because one of their targets is the cell cycle regulatory protein p27Kip1/CDKN1B [44]. In contrast to miR-221 and miR-222, different studies point to a tumor suppressor role for miR-503: in different cancer cell lines, it has been shown to target modulators of the cell cycle, such as cyclin-dependent kinases (CCND1, D2, D3) and cyclins (D1 and D3); consistently, miR-503 was found down-regulated in cancer tissues [reviewed in 45]. However, a possible differential contribution of miR-503 to tumors between males and females can presently not be evaluated since the gender context was ignored in current studies. This consideration is applicable also to other X-linked miRNAs implicated in different types of cancer [35].

With regard to breast cancer, the most common cancer in women, two profiling studies, performed on very large patient cohorts, indicate that circulating miRNAs from the two X-linked miR-106a-363 and miR-532-502 clusters are promising non-invasive biomarkers for diagnosis since they were upregulated in patients in comparison to healthy volunteers [46, 47]. Functional studies based on cell lines experiments showed an implication of some of those miRNA in breast cancer; in particular, miR-532, miR-19b and miR-20b act as oncomirs by down-regulating the tumor suppressors RERG (ras-related and estrogen-regulated growth inhibitor), PTPRG (protein tyrosine phosphatase receptor type G) and PTEN (phosphatase and tensin homologue), respectively [48–50]; in contrast, miR-188 and miR-502 act as tumor suppressors by inhibiting TMED3 (Transmembrane Emp24 Protein Transport Domain Containing 3) and the H4K20 methyltransferase SET8, respectively [51, 52]. Overall, further functional studies are required to put together the different pieces of the puzzle to gain a comprehensive view of possible X-linked miRNAs contribution to breast cancer.

Cardiovascular homeostasis

In the cardiovascular system, regulatory power of microR-NAs has been invoked at multiple level for controlling functions of various cells, such as cardiomyocytes, endothelial cells, smooth muscle cells, and fibroblasts [53]. Intriguingly, more women than man die of cardiovascular disease every year [54]. Based on the notion that estrogen can regulate transcription and miRNA processing, and that some X-linked miRNAs could escape XCI, it can be argued that miRNAs contribute to some sex-biased features of cardiovascular homeostasis [55]. Most of the studies into the field are based on profiling analyses of cellular and circulating miRNAs, with some insights into molecular mechanisms acquired from murine or rat models, whose extrapolation for humans should be viewed with caution. However, for some X-linked miRNAs, an association with biological pathways involved in cardiovascular homeostasis has been accomplished, although using human endothelial cells and thus requiring further validation in human subjects. For instance, miR-221/-222, in addition to their well-known role in cancer, are probably involved also in vascular and cardiac diseases since they can indirectly reduce the expression of endothelial nitric oxide synthase (eNOS), and NO impairment is a feature of different cardiomyopathies [53]; moreover, gender expression disparities have been reported, with decreased expression in at least murine hearts of miR-222, leading to higher levels of eNOS [56]. miR-92a and miR-223 inhibit angiogenesis by targeting integrin subunit alfa 5 and integrin beta 1, respectively [57, 58]. In contrast, miR-424 promote angiogenesis in human endothelial cells subjected to hypoxia by targeting cullin-2, a scaffolding protein critical to the assembly of the ubiquitin ligase system, thereby stabilizing the transcription factor hypoxia-inducible factor 1α (HIF- 1α), which regulates genes governing angiogenesis and metabolic pathways [59].

Male fertility

A biological pathway possibly regulated by X-linked miR-NAs that deserves investigation in the near future could be male fertility. In fact, it is recognized that the X chromosome is also correlated to male fertility, because of the presence of a relatively high number of testis-specific genes [60, 61]. This observation is probably applicable also to miR-NAs: Guo et al. demonstrated that the highest proportion of X-linked miRNAs was found in mouse testis; moreover, comparison between human and mouse showed that they had an average substitution rate higher than other testis



miRNAs on autosomes, implicating an miRNA contribution in male reproductive function and mammalian speciation [9, 62]. Supporting this hypothesis, one group of X-linked miRNAs was identified as a very quickly evolving miRNA cluster because of its presence restricted to primates; they are all hosted in the uncharacterized non-coding RNA (LOC105373347) (Fig. 1), and preferentially expressed in testicular tissues [63–65]. Functional investigations of the role of these miRNAs in male fertility are still missing. One miRNA from the cluster, miR-513, was shown to be able to downregulate the expression of the receptor for luteinizing hormone (LH) and human chorionic gonadotropin (hCG), two closely related hormones crucial not only for male but also female reproductive processes; however, the study was performed on primary granulosa cell cultures, thus requiring further validation in human subjects [66].

Targetome of X-linked miRNAs and its functional analysis using bioinformatics

Targetome and biological pathways

To possibly achieve new perspectives on the biological role of X-linked miRNAs, we decided to analyze their entire targetome and related biological pathways.

miRNAs exert their biological role by down-regulating specific targets; given that binding of miRNA to mRNA tolerates mismatches, except for the seed sequence (nucleotides 2–8 of the miRNA) requiring a perfect complementarity, each miRNA can potentially bind hundreds of mRNAs. Thus, identifying the entire "true" miRNA targetome is still challenging, even though highly informative concerning the role of miRNA in health and disease. Generally, the approach is based on a first step of bioinformatics prediction, followed by an experimental validation of those targets predicted by more than one bioinformatics tool. For our analyses, we considered 80 miRNAs directly related to the group of the 62 validated miRNAs, since some pre-miRNAs yield both -5p and -3p isoforms, experimentally validated, and in addition different pre-miRNAs produce the same mature miRNA (Supplementary Material). Exploring the miRWalk platform [67], for each miRNA we considered all the experimentally validated targets reported by the miRTarbase tool and predicted targets identified by both TargetScan and miRDB tools, with a cut-off score ≥ 0.5 for miRNA-mRNA pairings, in both UTRs and coding sequence of target transcripts. This approach produced a list of 6675 non-redundant transcripts, representing the targetome of X-linked miRNAs (Supplementary Material).

Pathway enrichment analysis of target genes was performed using the program DAVID (https://david.ncifcrf.gov/) that consists of an integrated biological

knowledgebase and analytic tools aimed at systematically extracting biological meaning from large gene/protein lists [68]. In particular, we focused on the significantly enriched pathways using KEGG (Fig. 2a and Supplementary Material) and Biocarta (Fig. 2b) as reference databases.

We found different pathways predicted by both tools, such as "p53 signaling pathway", "MAPK pathways" and particularly "Cell cycle" and its checkpoints. This last observation was already reported for enriched terms in the targetome of testis X-linked miRNAs and was put in relation to massive and continuous cell division during spermatogenesis [9]. Different pathways were also involved in immunity, and probably this is an expected result considering the data emerging from the literature and discussed in the previous paragraphs. KEGG pathways point also to involvement of the X-linked miRNA targetome in "Axon guidance" and "Neurotrophin pathway" (the second and the fourth most significantly enriched pathways, respectively); those fields are absolutely unexplored in terms of miRNA contribution, even though the contribution of genes on the X-chromosome to the nervous system development and functioning is known, as well as genes associated with X-linked intellectual disability (XLID) [69] and some X-linked miRNAs predicted to be involved in X-linked and autosomal intellectual disability (ID) gene regulation [70]. For example, HUWE1 is a gene implicated in XLID [71], and it hosts an miRNA cluster, whose possible contribution to the disease has not been explored yet.

The most significantly enriched pathway indicated by KEGG is "Pathways in cancer": this is probably not surprising, as microRNAs role in carcinogenesis represents one of the hot topic in contemporary research. However, specific contributions of X-linked miRNAs and/or gender context still need further study.

Finally, among the top-20-*p* value scoring of enriched pathways from KEGG emerges "Oocyte meiosis" and in the full list (Supplementary material) also "Progesteronemediated oocyte maturation": this field should be explored in the near future, also in terms of possible contribution to ovarian failure observed in Turner syndrome patients.

Methods

The list of 80 X-linked miRNAs was used to launch a search on miRWalk platform (http://mirwalk.umm.uni-heidelberg .de/) to retrieve all their target genes. In detail, by selecting "miRTarbase", or "TargetScan" and "miRDB", and "5' UTR", or "3' UTR" or "coding region" and selecting the lower score (0.5) for miRNA-mRNA pairings, 6 different target lists were retrieved containing all experimentally validated targets (by miRTarbase tool), or those predicted by both TargetScan and miRDB tools. By exporting the lists in Excel, a unique list of 6675 non-redundant transcripts



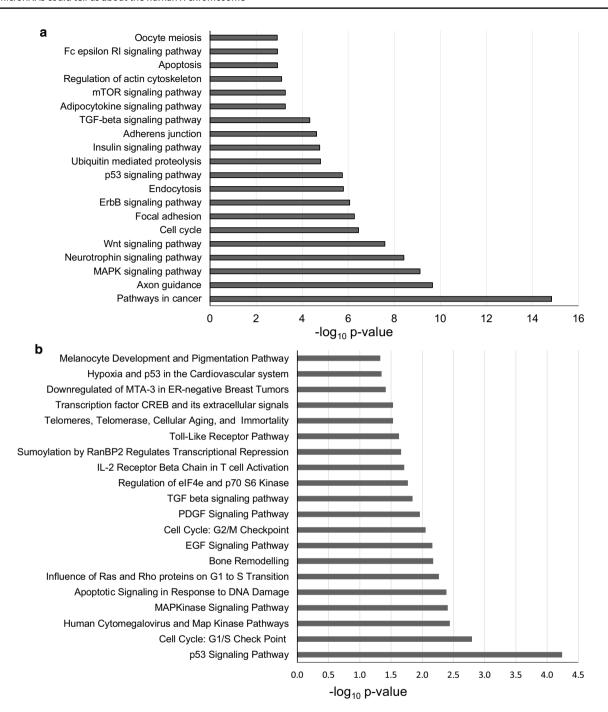


Fig. 2 Biological pathways potentially governed by X-linked microRNAs. The whole targetome (predicted and validated) of X-linked microRNAs was analyzed by the program DAVID. **a** Top-20-*p* values significantly enriched pathways according to KEGG (full list as Sup-

plementary Material) or **b** Biocarta. Biological pathways were considered statistically significant if p value was less than 0.05 (Benjamini–Hochberg procedure for multiple correction)

representing the whole targetome of X-linked miRNAs was produced. On that list, enrichment of biological pathways supplied by KEGG and Biocarta was performed by Database for Annotation, Visualization and Integrated Discovery (DAVID, http://david.abcc.ncifcrf.gov/), which facilitates the transition from data collection to biological analysis and

provides a comprehensive set of functional annotation tools to condense large gene lists into gene functional groups, to convert between gene/protein identifiers, to visualize manygene-to-many-term relationships, to cluster redundant and heterogeneous terms into groups, to search for interesting and related genes or terms, and, dynamically, to view genes



from their lists on bio-pathways. In detail, the pathway enrichment on our list of transcripts was quantitatively measured by statistical methods, including Chi-square, Fisher's exact test, binomial probability, and hypergeometric distribution. Considering this type of analysis could increase the false-positive rate, to control the false-positive rate in the results, a multiple test correction of enrichment p values was performed on the functional annotation categories by selecting the Benjamini–Hochberg procedure. Biological pathways with p values < 0.05 were considered statistically significant [72].

microRNAs in the genotype-phenotype relationship of Turner and Klinefelter syndrome patients

The partial or complete lack of a second X chromosome in a female causes Turner syndrome, estimated 1 in 2000 females [73]. 40-50% of Turner Syndrome women have the 45,X karyotype, 15-25% have mosaicism with 45,X/46,XX and about 3% present with 45,X/46,XY. 20% of cases have alterations of the X chromosome, with isochromosome Xq being the most frequent; ring X chromosome with deletion of either Xp or Xq are also present and 10–12% of cases have differing amounts of Y chromosome material. Phenotypic traits include short stature, infertility, cardiac malformations, bone dysgenesis, neurocognitive challenges, and autoimmune diseases, but also some external features such as short neck with webbing, and lymphedema; however, the external appearance and internal anomalies of Turner syndrome individuals vary greatly and cannot be predicted based on the karyotype—thus, there does not seem to be a clear karyotype-phenotype relation. When diagnosed in the childhood, Turner syndrome patients are offered therapy with growth hormone aimed at increasing adult height and later with sex hormone replacement therapy, due to gonadal dysgenesis resulting in hypergonadotropic hypogonadism [74].

It has become evident that Turner syndrome is much more than simple genetics, as no obvious karyotype—phenotype relationship exists: two individuals with the exact same karyotypes, can exhibit very different traits and comorbidities. The relationship between karyotype and phenotype is certainly complicated by the mosaicism (presence of more than one cell lines with X aneuploidies), that varies with tissue type and age of a given person, but also suggests a role for epigenetics mechanisms [75]. So far, few studies have been dedicated to that issue, essentially based on analysis of DNA methylation and histone modifications [16]. Regarding miRNAs, one paper unveils the role of miR-320a, found upregulated in PBMC and fetal gonadal tissues of 45,X TS females, and thus causing downregulation of its target

KITLG, essential for ovarian development and primordial cell survival. This mechanism could partially account for the Turner syndrome-associated gonadal dysgenesis. miR-320 is not an X-linked miRNA, but its expression is modulated by *KDM5C*, encoded by an XCI-escaping gene [18], which is also differentially expressed in Turner syndrome [14]. Furthermore, a number of circulating miRNAs were differentially detected in 45,X Turner syndrome females as compared with 46,XX normal women, whose role should be further clarified [19].

Presence of an extra X-chromosome characterizes Klinefelter syndrome males, 1 in every 660 male births. Klinefelter syndrome is characterized by a 47,XXY karyotype in about 80-90% of men, whereas the remaining cases are represented by mosaicism, additional sex chromosome (e.g., 48,XXXY; 48,XXYY; 49,XXXXY) or X chromosome structural abnormalities. The low testosterone levels, hypergonadotropic hypogonadism observed in a high proportion of these men only explain a few of the classical characteristics of Klinefelter syndrome, such as tall stature, gynecomastia, while the frequent infertility encountered is a consequence of the dramatic demise of the seminiferous tubules which takes place early in life [76]. Also in the case of Klinefelter syndrome, the wide variable phenotypic spectrum and the different severity of symptoms suggest a role for epigenetic factors. Two different studies have profiled miRNAs in peripheral blood cells from a small group of non-mosaic Klinefelter syndrome patients and healthy men and found 89 [20] or 2 [21] miRNAs differentially detectable. Also in the case of Klinefelter syndrome, the functional contribution of miRNAs to the phenotype has not been explored.

Potential key X-linked genes responsible for the comorbidities seen in Turner syndrome, Klinefelter syndrome and other X chromosome aneuploidy syndromes have been identified. One example is represented by KDM6A, involved in germ cell development, differentially expressed, and methylated in both syndromes [14]; another example is KDM5C, which could play a role in the neurocognitive development of the syndromes [77]; TIMP1/TIMP3 genes are involved in the bicuspid aortic valve, probably accounting for higher incidence of aortic dissection in Turner syndrome [78]. The SHOX gene (short stature homeobox), located on Xp22.23, is the only X-chromosome gene that is truly linked with a phenotypic trait in Turner syndrome, i.e., short stature and skeletal growth [79]. The function is dosage dependent, thus causing short stature in Turner syndrome and increased stature in other X aneuploidy condition characterized by extra X chromosomes (47,XXX; 47,XXY; 48,XXYY) [80].

With regard to miRNAs, currently only speculations are possible. For example, taking into account the number of X-linked miRNAs with a role in immunity, it is paradigmatic that a plethora of autoimmunity stigmata are observed in Turner syndrome patients [81]; consistently, Klinefelter



syndrome males (47,XXY) have a risk similar to female to develop not only systemic lupus erythematosus (SLE), an autoimmune disease with a striking female preponderance [82], but also other autoimmune conditions [83] and as such both missing an X chromosome (like in Turner syndrome) and having too many (like in Klinefelter syndrome) can lead to an increased risk of autoimmune disease. The number of X chromosomes, and the consequent altered X-linked genes and miRNAs dosage, is probably critical for the maintenance or the loss of the immune homeostasis. Again, a role for X-linked miRNA in infertility of Klinefelter syndrome men could be hypothesized, based on some data in the literature discussed above: miRNAs are essential for male fertility, as shown by spermatogonial knock-out of their processing enzyme Dicer [61, 84]; the highest proportion of X-linked miRNAs is observed in mouse testis; 86% of the X-linked miRNAs escape from X inactivation during meiosis and thus might be crucial for gene expression regulation during spermatogenesis [85]. Finally, we inquired also if the targetome of X-linked miRNAs was already known to be involved in diseases using OMIM as reference database (DAVID search): five out eight statistically significant items were "diabetes"; consistently, "Insulin signaling pathway" is one of significantly enriched pathways according to KEGG. These bioinformatics results should be experimentally investigated, taking into account that diabetes is seen with four to sixfold increased frequency in Turner syndrome and Klinefelter syndrome adult patients [75, 86].

Overall, focusing the research on miRNA contribution to Turner syndrome and Klinefelter syndrome phenotype would take the understanding of syndrome development and associated morbidities to a new level, likely also in terms of therapeutic approach. For example, systematic studies on a correlation between the absence or extra-copies of specific X regions containing miRNAs, with consequent dysregulation of their potential targets, and specific clinical features should be performed. Furthermore, miRNA profiling on different tissues other than blood cells could prompt functional analysis on those miRNAs differentially expressed between X-aneuploidy and normal individuals. These studies could also pave the way for the development of new therapeutic approaches (e.g., co-administration of GH or sex hormone with an "essential" synthetic miRNA in TS).

Conclusion and perspectives

Females appear to be equipped with a larger miRNA machinery than males: X chromosome contains an unexpected high number of miRNAs (118), in comparison to Y chromosome (4 miRNA sequences predicted) and an average of 40–50 localized on autosomes. Taking into consideration the regulatory power of these small non-coding RNAs, the scenario

depicted above suggests a strong miRNA involvement in sex-specific phenotypes and some differences in pathogenetic mechanisms and/or pathological responses. This is also supported by the fact that also at cellular level, differences in response to stressful stimuli have been observed between cells carrying XX or XY chromosomes, with a possible direct role of miRNAs; as an example, miR-548am has been shown to act as a key modulator of sex difference in the susceptibility to mitochondria-based apoptosis in primary dermal fibroblasts [29]. However, most of the miRNA-based studies ignore the gender context. Some sex-biased expression of specific miRNAs could account for sex different pathophysiological conditions. This research field should be explored in the next future, to fill the gap between clinical data and our understanding of molecular mechanisms underlying some gender-biased diseases, also in terms of possible new therapeutic approaches. Noteworthy, to date most of the X-linked miRNAs have no described functions. In addition, in this review, only X-linked validated miRNAs have been considered, but they represent approximately the half of miRNA content of X chromosome; all the other miRNAs should be validated to have a comprehensive picture of their contribution to X chromosome properties and implication in the pathogenesis of X aneuploidy syndromes.

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References

- Ambros V (2004) The functions of animal microRNAs. Nature 431:350–355. https://doi.org/10.1038/nature02871
- Lewis BP, Burge CB, Bartel DP (2005) Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. Cell 120(1):15–20. https://doi. org/10.1016/j.cell.2004.12.035
- Filipowicz W, Bhattacharyya SN, Sonenberg N (2008) Mechanisms of post-transcriptional regulation by microRNAs: are the answers in sight? Nat Rev Genet 9(2):102–114. https://doi.org/10.1038/nrg2290
- Chang TC, Mendell JT (2007) microRNAs in vertebrate physiology and human disease. Annu Rev Genom Hum Genet 8:215–239. https://doi.org/10.1146/annurev.genom.8.080706.092351
- Di Leva G, Garofalo M, Croce CM (2014) MicroRNAs in cancer. Annu Rev Pathol 9:287–314. https://doi.org/10.1146/annurev-pathol-012513-104715
- Potenza N, Papa U, Scaruffi P, Mosca N, Tonini GP, Russo A (2010) A novel splice variant of the human dicer gene is expressed in neuroblastoma cells. FEBS Lett 584(15):3452–3457. https://doi.org/10.1016/j.febslet.2010.06.045



- Olena AF, Patton JG (2010) Genomic organization of microRNAs. J Cell Physiol 222:540–545. https://doi.org/10.1002/jcp.21993
- Bartel DP (2004) MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 116(2):281–297. https://doi.org/10.1016/s0092 -8674(04)00045-5
- Guo X, Su B, Zhou Z, Sha J (2009) Rapid evolution of mammalian X-linked testis microRNAs. BMC Genom 10:97. https://doi. org/10.1186/1471-2164-10-97
- Lyon MF (1961) Gene action in the X-chromosome of the mouse (*Mus musculus* L.). Nature 190:372–373. https://doi. org/10.1038/190372a0
- Carrel L, Willard HF (2005) X-inactivation profile reveals extensive variability in X-linked gene expression in females. Nature 434(7031):400–404. https://doi.org/10.1038/nature03479
- Brooks WH (2010) X chromosome inactivation and autoimmunity. Clin Rev Allergy Immunol 39(1):20–29. https://doi.org/10.1007/s12016-009-8167-5
- Disteche CM, Berletch JB (2015) X-chromosome inactivation and escape. J Genet 94(4):591–599. https://doi.org/10.1007/s1204 1-015-0574-1
- Trolle C, Nielsen MM, Skakkebæk A, Lamy P, Vang S, Hedegaard J, Nordentoft I, Ørntoft TF, Pedersen JS, Gravholt CH (2016) Widespread DNA hypomethylation and differential gene expression in Turner syndrome. Sci Rep 6:34220. https://doi.org/10.1038/srep34220
- Sharma A, Jamil MA, Nuesgen N, Schreiner F, Priebe L, Hoffmann P, Herns S, Nöthen MM, Fröhlich H, Oldenburg J, Woelfle J, El-Maarri O (2015) DNA methylation signature in peripheral blood reveals distinct characteristics of human X chromosome numerical aberrations. Clin Epigenet. https://doi.org/10.1186/s13148-015-0112-2
- Viuff M, Skakkebaek A, Nielsen MN, Chang S, Gravholt CH (2019) Epigenetics and genomics in Turner Syndrome. Am J Med Genet C Semin Med Genet 181(1):68–75. https://doi.org/10.1002/ ajmg.c.31683
- Álvarez-Nava F, Lanes R (2018) Epigenetics in Turner syndrome. Clin Epigenet 10:45. https://doi.org/10.1186/s13148-018-0477-0
- Zhang X, Hong D, Ma S, Ward T, Ho M, Pattni R, Duren Z, Stankov A, Bade Shrestha S, Hallmayer J, Wong WH, Reiss AL, Urban AE (2020) Integrated functional genomic analyses of Klinefelter and Turner syndromes reveal global network effects of altered X chromosome dosage. Proc Natl Acad Sci USA 117(9):4864–4873. https://doi.org/10.1073/pnas.1910003117
- Sun YX, Zhang YX, Zhang D, Xu CM, Chen SC, Zhang JY, Ruan YC, Chen F, Zhang RJ, Qian YQ, Liu YF, Jin LY, Yu TT, Xu HY, Luo YQ, Liu XM, Sun F, Sheng JZ, Huang FHF (2017) XCI-escaping gene KDM5C contributes to ovarian development via downregulating miR-320a. Hum Genet 136(2):227–239. https://doi.org/10.1007/s00439-016-1752-9
- Sui W, Ou M, Chen J, Li H, Lin H, Zhang Y, Li W, Xue W, Tang D, Gong W, Zhang R, Li F, Dai Y (2012) microRNA expression profile of peripheral blood mononuclear cells of Klinefelter Syndrome. Exp Ther Med 4(5):825–831. https://doi.org/10.3892/etm.2012.682
- Cimino L, Salemi M, Cannarella R, Condorelli RA, Giurato G, Marchese G, La Vignera S, Calogero AE (2017) Decreased miRNA expression in Klinefelter Syndrome. Sci Rep 7(1):16672. https://doi.org/10.1038/s41598-017-16892-3
- Lahn BT, Page DC (1999) Four evolutionary strata on the human X chromosome. Science 286(5441):964–967. https://doi. org/10.1126/science.286.5441.964
- 23. Santos-Rebouças CB, de Almeida LG, Belet S, Dos Santos SR, Ribeiro MG, da Silva AF, Medina-Acosta E, Dos Santos JM, Gonçalves AP, Bahia PR, Pimentel MM, Froyen G (2015) Novel microduplications at Xp11.22 including HUWE1: clinical and molecular insights into these genomic rearrangements associated

- with intellectual disability. J Hum Genet 4:207–211. https://doi.org/10.1038/jhg.2015.1
- Prakash SK, Bondy CA, Maslen CL, Silberbach M, Lin AE, Perrone L, Limongelli G, Michelena HI, Bossone E, Citro R, Citro R, Lemaire SA, Milewicz DM, BAVCon Investigators, GenTAC Registry Investigators (2016) Autosomal and X chromosome structural variants are associated with congenital heart defects in Turner syndrome: the NHLBI GenTAC registry. Am J Med Genet A 170(12):3157–3164. https://doi.org/10.1002/ajmg.a.37953
- Shen L, Liu C, Gao M, Li H, Zhang Y, Tian Q, Ni H, Peng P, Zhao R, Hu Z, Gao Y, Xia K, Bo Q, Guo H (2019) Novel mutation of EDA causes new asymmetrical X-linked hypohidrotic ectodermal dysplasia phenotypes in a female. J Dermatol 46(8):731–733. https://doi.org/10.1111/1346-8138.14978
- Tsang SH, Sharma T (2018) X-linked choroideremia. Adv Exp Med Biol 1085:37–42. https://doi.org/10.1007/978-3-319-95046 -4 9
- Chureau C, Chantalat S, Romito A, Galvani A, Duret L, Avner P, Rougeulle C (2011) Ftx is a non-coding RNA which affects Xist expression and chromatin structure within the X-inactivation center region. Hum Mol Genet 20(4):705–718. https://doi.org/10.1093/hmg/ddq516
- Liu F, Yuan JH, Yang F, Wang TT, Ma JZ, Zhang L, Zhou CC, Wang F, Yu J, Zhou WP, Sun SH (2016) Long noncoding RNA FTX inhibits hepatocellular carcinoma proliferation and metastasis by binding MCM2 and miR-374a. Oncogene 35(41):5422-5434. https://doi.org/10.1038/onc.2016.80
- Matarrese P, Tieri P, Anticoli S, Ascione B, Conte M, Franceschi C, Malorni W, Salvioli S, Ruggieri A (2019) X-chromosomelinked miR548am-5p is a key regulator of sex disparity in the susceptibility to mitochondria-mediated apoptosis. Cell Death Dis 10(11):828. https://doi.org/10.1038/s41419-019-2069-0
- Klein SL, Flanagan KL (2016) Sex differences in immune responses. Nat Rev Immunol 16(10):626–638. https://doi. org/10.1038/nri.2016.90
- Carè A, Bellenghi M, Matarrese P, Gabriele L, Salvioli S, Malorni W (2018) Sex disparity in cancer: roles of microRNAs and related functional players. Cell Death Differ 25(3):477–485. https://doi.org/10.1038/s41418-017-0051-x
- Fairweather DL, Frisancho-Kiss S, Rose NR (2008) Sex differences in autoimmune disease from a pathological perspective.
 Am J Pathol 173(3):600–609. https://doi.org/10.2353/ajpat h.2008.071008
- Libert C, Dejager L, Pinheiro I (2010) The X chromosome in immune functions: when a chromosome makes the difference. Nat Rev Immunol 10(8):594–604. https://doi.org/10.1038/nri2815
- Bianchi I, Lleo A, Gershwin ME, Invernizzi P (2012) The X chromosome and immune associated genes. J Autoimmun 38(2–3):J187–J192. https://doi.org/10.1016/j.jaut.2011.11.012
- Pinheiro I, Dejager L, Libert C (2011) X-chromosome-located microRNAs in immunity: might they explain male/female differences? The X chromosome-genomic context may affect X-located miRNAs and downstream signaling, thereby contributing to the enhanced immune response of females. BioEssays 33(11):791– 802. https://doi.org/10.1002/bies.201100047
- Lindsay MA (2008) microRNAs and the immune response. Trends Immunol 29(7):343–351. https://doi.org/10.1016/j.it.2008.04.004
- Johnnidis JB, Harris MH, Wheeler RT, Stehling-Sun S, Lam MH, Kirak O, Brummelkamp TR, Fleming MD, Camargo FD (2008) Regulation of progenitor cell proliferation and granulocyte function by microRNA-223. Nature 451(7182):1125–1129. https://doi. org/10.1038/nature06607
- Landais S, Landry S, Legault P, Rassart E (2007) Oncogenic potential of the miR-106–363 cluster and its implication in human T-cell leukemia cancer. Cancer Res 67(12):5699–5707. https:// doi.org/10.1158/0008-5472.CAN-06-4478



- Forrest AR, Kanamori-Katayama M, Tomaru Y, Lassmann T, Ninomiya N, Takahashi Y, de Hoon MJL, Kubosaki A, Kaiho A, Suzuki M, Yasuda J, Kawai J, Hayashizaki Y, Hume DA, Suzuki H (2010) Induction of microRNAs, mir-155, mir-222, mir-424 and mir-503, promotes monocytic differentiation through combinatorial regulation. Leukemia 24(2):460–466. https://doi.org/10.1038/ leu.2009.246
- Khalifa O, Pers YM, Ferreira R, Sénéchal A, Jorgensen C, Apparailly F, Duroux-Richard I (2016) X-linked miRNAs associated with gender differences in rheumatoid arthritis. Int J Mol Sci 17(11):1852. https://doi.org/10.3390/ijms17111852
- Mokdad AH, Dwyer-Lindgren L, Fitzmaurice C, Stubbs RW, Bertozzi-Villa A, Morozoff C, Charara R, Allen C, Naghavi M, Murray CJ (2017) Trends and patterns of disparities in cancer mortality among US counties, 1980–2014. JAMA 317(4):388– 406. https://doi.org/10.1001/jama.2016.20324
- Garofalo M, Quintavalle C, Romano G, Croce CM, Condorelli G (2012) miR221/222 in cancer: their role in tumor progression and response to therapy. Curr Mol Med 12(1):27–33. https://doi. org/10.2174/156652412798376170
- 43. Di Leva G, Gasparini P, Piovan C, Ngankeu A, Garofalo M, Taccioli C, Iorio MV, Li M, Volinia S, Alder H, Nakamura T, Nuovo G, Liu Y, Nephew KP, Croce CM (2010) MicroRNA cluster 221–222 and estrogen receptor alpha interactions in breast cancer. J Natl Cancer Inst 102(10):706–721. https://doi.org/10.1093/jnci/djq10
- 44. le Sage C, Nagel R, Egan DA, Schrier M, Mesman E, Mangiola A, Anile C, Maira G, Mercatelli N, Ciafrè SA, Farace MG, Agami R (2007) Regulation of the p27(Kip1) tumor suppressor by miR-221 and miR-222 promotes cancer cell proliferation. EMBO J 26(15):3699–3708. https://doi.org/10.1038/sj.emboj.7601790
- Gupta G, Chellappan DK, de Jesus Andreoli Pinto T, Hansbro PM, Bebawy M, Dua K (2018) Tumor suppressor role of miR-503.
 Panminerva Med 60(1):17–24. https://doi.org/10.23736/S0031-0808.17.03386-9
- 46. Li M, Zhou Y, Xia T, Zhou X, Huang Z, Zhang H, Zhu W, Ding Q, Wang S (2018) Circulating microRNAs from the miR-106a-363 cluster on chromosome X as novel diagnostic biomarkers for breast cancer. Breast Cancer Res Treat 170(2):257–270. https://doi.org/10.1007/s10549-018-4757-3
- Zhou X, Liu Q, Xia T, Zhu W (2020) Circulating miR-532–502 cluster derived from chromosome X as biomarkers for diagnosis of breast cancer. Gene 722:144104. https://doi.org/10.1016/j.gene.2019.144104
- Huang L, Tang X, Shi X, Su L (2020) miR-532-5p promotes breast cancer proliferation and migration by targeting RERG. Exp Ther Med 19(1):400–408. https://doi.org/10.3892/etm.2019.8186
- Liu M, Yang R, Urrehman U, Ye C, Yan X, Cui S, Hong Y, Gu Y, Liu Y, Zhao C, Yan L, Zhang CY, Liang H, Chen X (2016) MiR-19b suppresses PTPRG to promote breast tumorigenesis. Oncotarget 7(39):64100–64108. https://doi.org/10.18632/oncotarget.11799
- Zhou W, Shi G, Zhang Q, Wu Q, Li B, Zhang Z (2014) Micro-RNA-20b promotes cell growth of breast cancer cells partly via targeting phosphatase and tensin homologue (PTEN). Cell Biosci 4(1):62. https://doi.org/10.1186/2045-3701-4-62
- Pei J, Zhang J, Yang X, Wu Z, Sun C, Wang Z, Wang B (2019) TMED3 promotes cell proliferation and motility in breast cancer and is negatively modulated by miR-188-3p. Cancer Cell Int. 19:75. https://doi.org/10.1186/s12935-019-0791-4
- 52. Liu B, Zhang X, Song F, Zheng H, Zhao Y, Li H, Zhang L, Yang M, Zhang W, Chen K (2016) MiR-502/SET8 regulatory circuit in pathobiology of breast cancer. Cancer Lett 376(2):259–267. https://doi.org/10.1016/j.canlet.2016.04.008

- Urbich C, Kuehbacher A, Dimmeler S (2008) Role of microRNAs in vascular diseases, inflammation, and angiogenesis. Cardiovasc Res 79(4):581–588. https://doi.org/10.1093/cvr/cvn156
- Garcia M, Mulvagh SL, Merz CN, Buring JE, Manson JE (2016) Cardiovascular disease in women: clinical perspectives. Circ Res 118(8):1273–1293. https://doi.org/10.1161/CIRCRESAHA .116.307547
- 55. Florijn BW, Bijkerk R, van der Veer EP, van Zonneveld AJ (2018) Gender and cardiovascular disease: are sex-biased microRNA networks a driving force behind heart failure with preserved ejection fraction in women? Cardiovasc Res 114(2):210–225. https://doi.org/10.1093/cyr/cyx223
- Evangelista AM, Deschamps AM, Liu D, Raghavachari N, Murphy E (2013) miR-222 contributes to sex-dimorphic cardiac eNOS expression via ets-1. Physiol Genom 45(12):493–498. https://doi.org/10.1152/physiolgenomics.00008.2013
- Bonauer A, Carmona G, Iwasaki M, Mione M, Koyanagi M, Fischer A, Burchfield J, Fox H, Doebele C, Ohtani K, Chavakis E, Potente M, Tjwa M, Urbich C, Zeiher AM, Dimmeler S (2009) MicroRNA-92a controls angiogenesis and functional recovery of ischemic tissues in mice. Science 324(5935):1710–1713. https://doi.org/10.1126/science.1174381
- Shi L, Fisslthaler B, Zippel N, Frömel T, Hu J, Elgheznawy A, Heide H, Popp R, Fleming I (2013) MicroRNA-223 antagonizes angiogenesis by targeting β1 integrin and preventing growth factor signaling in endothelial cells. Circ Res 113(12):1320–1330. https://doi.org/10.1161/CIRCRESAHA.113.301824
- Ghosh G, Subramanian IV, Adhikari N, Zhang X, Joshi HP, Basi D, Chandrashekhar YS, Hall JL, Roy S, Zeng Y, Ramakrishnan S (2010) Hypoxia-induced microRNA-424 expression in human endothelial cells regulates HIF-α isoforms and promotes angiogenesis. J Clin Investig 120(11):4141–4154. https://doi.org/10.1161/10.1172/JCI42980
- Stouffs K, Lissens W (2012) X chromosomal mutations and spermatogenic failure. Biochim Biophys Acta 1822(12):1864–1872. https://doi.org/10.1016/j.bbadis.2012.05.012
- Papaioannou MD, Nef S (2010) microRNAs in the testis: building up male fertility. J Androl 31(1):26–33. https://doi.org/10.2164/jandrol.109.008128
- Wang PJ, McCarrey JR, Yang F, Page DC (2001) An abundance of X-linked genes expressed in spermatogonia. Nat Genet 27(4):422– 426. https://doi.org/10.1038/86927
- Bentwich I, Avniel A, Karov Y, Aharonov R, Gilad S, Barad O, Barzilai A, Einat P, Einav U, Meiri E, Sharon E, Spector Y, Bentwich Z (2005) Identification of hundreds of conserved and non-conserved human microRNAs. Nat Genet 37(7):766–770. https://doi.org/10.1038/ng1590
- 64. Zhang R, Peng Y, Wang W, Su B (2007) Rapid evolution of an X-linked microRNA cluster in primates. Genome Res 17(5):612– 617. https://doi.org/10.1101/gr.6146507
- 65. Ramaiah M, Tan K, Plank TD, Song M, Dumdie JN, Jones S, Shum EY, Sheridan SD, Peterson KJ, Gromoll J, Haggarty SJ, Cook-Andersen H, Wilkinson MF (2019) A microRNA cluster in the fragile-X region expressed during spermatogenesis targets FMR1. EMBO Rep 20(2):e46566
- Troppmann B, Kossack N, Nordhoff V, Schüring AN, Gromoll J (2014) MicroRNA miR-513a-3p acts as a co-regulator of luteinizing hormone/chorionic gonadotropin receptor gene expression in human granulosa cells. Mol Cell Endocrinol 390(1–2):65–72. https://doi.org/10.1016/j.mce.2014.04.003
- Sticht C, De La Torre C, Parveen A, Gretz N (2018) miRWalk: an online resource for prediction of microRNA binding sites. PLoS One. https://doi.org/10.1371/journal.pone.0206239
- Huang DW, Sherman BT, Lempicki RA (2009) Systematic and integrative analysis of large gene lists using DAVID



- bioinformatics resources. Nat Protoc 4(1):44–57. https://doi.org/10.1038/nprot.2008.211
- Stevenson RE, Schwartz CE, Rogers RC (2012) Atlas of X-linked intellectual disability syndromes, 2nd edn. Oxford University Press, New York
- Gonçalves TF, Piergiorge RM, Dos Santos JM, Gusmão J, Pimentel MMG, Santos-Rebouças CB (2019) Network profiling of brain-expressed X-chromosomal microRNA genes implicates shared key microRNAs in intellectual disability. J Mol Neurosci 67(2):295–304. https://doi.org/10.1007/s12031-018-1235-7
- Froyen G, Belet S, Martinez F, Santos-Rebouças CB, Declercq M, Verbeeck J, Donckers L, Berland S, Mayo S, Rosello M, Pimentel MM, Fintelman-Rodrigues N, Hovland R, Rodrigues dos Santos S, Raymond FL, Bose T, Corbett MA, Sheffield L, van Ravenswaaij-Arts CM, Dijkhuizen T, Coutton C, Satre V, Siu V, Marynen P (2012) Copy-number gains of HUWE1 due to replication- and recombination-based rearrangements. Am J Hum Genet 91(2):252–264. https://doi.org/10.1016/j.ajhg.2012.06.010
- 72. Huang DW, Sherman BT, Lempicki RA (2009) Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. Nucleic Acids Res 37(1):1–13. https://doi.org/10.1093/nar/gkn923
- Berglund A, Viuff MH, Skakkebæk A, Chang S, Stochholm K, Gravholt CH (2019) Changes in the cohort composition of turner syndrome and severe non-diagnosis of Klinefelter, 47, XXX and 47, XYY syndrome: a nationwide cohort study. Orphanet J Rare Dis 14(1):16. https://doi.org/10.1186/s13023-018-0976-2
- 74. Lin AE, Prakash SH, Andersen NH, Viuff M, Levitsky LL, Rivera-Davila M, Crenshaw ML, Hansen L, Colvin MK, Hayes FJ, Lilly E, Snyder EA, Nader-Eftekhari S, Aldrich MB, Bhatt AB, Prager LM, Arenivas A, Skakkebaek A, Steeves MA, Kreher JB, Gravholt CH (2019) Recognition and management of adults with Turner syndrome: from the transition of adolescence through the senior years. Am J Med Genet A 179(10):1987–2033. https://doi.org/10.1002/ajmg.a.61310
- Gravholt CH, Viuff MH, Brun S, Stochholm K, Andersen NH (2019) Turner syndrome: mechanisms and management. Nat Rev Endocrinol 15(10):601–614. https://doi.org/10.1038/s4157 4-019-0224-4
- Aksglaede L, Juul A (2013) Testicular function and fertility in men with Klinefelter syndrome: a review. Eur J Endocrinol 168(4):R67–R76. https://doi.org/10.1530/EJE-12-0934
- Skakkebæk A, Nielsen MM, Trolle C, Vang S, Hornshøj H, Hedegaard J, Wallentin M, Bojesen A, Hertz JM, Fedder J, Østergaard JR, Pedersen JS, Gravholt CH (2018) DNA hypermethylation and differential gene expression associated with Klinefelter syndrome. Sci Rep 8(1):13740. https://doi.org/10.1038/s41598-018-31780-0
- 78. Corbitt H, Morris SA, Gravholt CH, Mortensen KH, Tippner-Hedges R, Silberbach M, Maslen CL, Investigators GenTAC

- Registry (2018) TIMP3 and TIMP1 are risk genes for bicuspid aortic valve and aortopathy in Turner syndrome. PLoS Genet 14(10):e1007692. https://doi.org/10.1371/journal.pgen.1007692
- 79. Rao E, Weiss B, Fukami M, Rump A, Niesler B, Mertz A, Muroya K, Binder G, Kirsch S, Winkelmann M, Nordsiek G, Heinrich U, Breuning MH, Ranke MB, Rosenthal A, Ogata T, Rappold GA (1997) Pseudoautosomal deletions encompassing a novel homeobox gene cause growth failure in idiopathic short stature and Turner syndrome. Nat Genet 16(1):54–63. https://doi.org/10.1038/ng0597-54
- Ottesen AM, Aksglaede L, Garn I, Tartaglia N, Tassone F, Gravholt CH, Bojesen A, Sørensen K, Jørgensen N, Rajpert-De Meyts E, Gerdes T, Lind AM, Kjaergaard S, Juul A (2010) Increased number of sex chromosomes affects height in a nonlinear fashion: a study of 305 patients with sex chromosome aneuploidy. Am J Med Genet A 152A(5):1206–1212. https://doi. org/10.1002/ajmg.a.33334
- 81. Salerno M, Capalbo D, Cerbone M, De Luca F (2016) Subclinical hypothyroidism in childhood—current knowledge and open issues. Nat Rev Endocrinol 12(12):734–746. https://doi. org/10.1038/nrendo.2016.100
- 82. Sawalha AH, Harley JB, Scofield RH (2009) Autoimmunity and Klinefelter's syndrome: when men have two X chromosomes. J Autoimmun 33(1):31–34. https://doi.org/10.1016/j.jaut.2009.03.006
- Bojesen A, Juul S, Birkebaek NH, Gravholt CH (2006) Morbidity in Klinefelter syndrome: a Danish register study based on hospital discharge diagnoses. J Clin Endocrinol Metab 91(4):1254–1260. https://doi.org/10.1210/ic.2005-0697
- Korhonen HM, Meikar O, Yadav RP, Papaioannou MD, Romero Y, Da Ros M, Herrera PL, Toppari J, Nef S, Kotaja N (2011) Dicer is required for haploid male germ cell differentiation in mice. PLoS One 6(9):e24821. https://doi.org/10.1371/journ al.pone.0024821
- Song R, Ro S, Michaels JD, Park C, McCarrey JR, Yan W (2009) Many X-linked microRNAs escape meiotic sex chromosome inactivation. Nat Genet 41(4):488–493. https://doi.org/10.1038/ng.338
- Gravholt CH, Chang S, Wallentin M, Fedder J, Moore P, Skakkebæk A (2018) Klinefelter Syndrome: integrating genetics, neuropsychology, and endocrinology. Endocr Rev 39(4):389–423. https://doi.org/10.1210/er.2017-00212

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